

Molecular characterization of *Rhipicephalus microplus* using sangersequencing COI&18SrRNA Markersfrom Chhatrapati Sambhajinagar, India

Abstract

Genus *Rhipicephalus* ticks, which infest cattle, are the main animal parasites that cause billions of rupees' worth of annual economic losses. Species identification is highly challenging because of the physical similarities among the members of the *Rhipicephalus* (*Boophilus*) genus. The adult of *R. microplus* ticks from Chh. Sambhajinagar (MS) India, were examined morphologically and molecularly in this work. The morphology of the *R. microplus* isolates was different from that of the real *R. microplus* clade A ticks. Out of 13 sample of species 2 species analysed and successful sequencing outcome aby using sanger sequencing COI&18SrRNA marker the database inserted into BLASTncbi database and result matches with *Rhipicephalus microplus*. Our study looked into tick diversity using a molecular technique that included phylogenetic analysis, DNA sequencing, and PCR.

Keywords: *Rhipicephalus microplus*, Sanger sequencing, COI, 18s rRNA.

Introduction

Parasitism is relationship between species where one organism the parasite, live on or inside another organism. The host causing it some harm and its adapted structurally to this way of life. parasitic organism reduces host fitness. Ectoparasites are mostly arthropods found in cattle that either cause diseases or act as vector transmitting other parasites and endoparasites are classified into intestinal, atrial may inhabit body tissue causing serious health problems (Manar M.S. EI-Tons, introduction to medical parasitology). From ancient world the cattle industry is one of the most important and profitable agribusiness activities in the world. Livestock play an important role in Indian economy near about 20.5 million people depends upon livestock for their live hood (Livestock census 2020) parasites cause negative impacts parasite have on health and welfare of animal can include Blood loss if Substantial can lead to anemia and death, reducing growth rate, reducing reproductive rate, reducing milk production.

“In Chh. Sambhajinagar district rural area depends upon livestock therefore aim to purpose of research to collect and identify, parasites on Cattles. In many parts of the world *Rhipicephalus* (*Boophilus*) *microplus* is known simply as the cattle tick. This is specially so where it has spread from its origin in south east Asia to major cattle ranching areas like South America. This spread has been accidental, on commercial cattle transportations. In Africa this tick has established in much of southern and eastern Africa and it is widespread in Madagascar. *Rhipicephalus* (*Bo.*) *microplus* is more dangerous than *Rh. (Boophilus) decoloratus* because it transmits both *Babesia bovis* and *Babesia bigemina*, in comparison to *Rh. (Bo.) decoloratus* which transmits only *Ba. bigemina*, the less pathogenic of the two protozoans. For this reason the ability to identify *Rh. (Bo.) microplus* is most important as an aid to prevention of the spread of *Ba. bovis*. This tick

species and the others in the *Boophilus* sub-genus within the genus *Rhipicephalus* are so well known in their original classification as members of the full genus *Boophilus* that the former names are likely to remain in use for many years without confusion. The character states that have been used for this sub-genus are separate from the rest of the genus *Rhipicephalus*” (Tamura et al., 2021).

“*Rhipicephalus (Bo.) microplus* is a one-host tick with a monotropic type of behaviour. The time spent by the three stages on the host is about three weeks and egg laying can be completed in about four weeks. This is faster than *Rh. (Bo.) decoloratus*, moreover female *Rh. (Bo.) microplus* lay approximately 500 eggs more than *Rh. (B.) decoloratus* females. The steady spread of *Rh. (Bo.) microplus* in Africa is assisted by this higher reproductive potential which enables it to compete successfully against *Rh. (Bo.) decoloratus* where these ticks occur together in climates that are most favourable to *Rh. (Bo.) microplus*. Large numbers of larvae are usually present on the vegetation in late spring, and successive generations of larvae then occur through the summer and into the cooler autumn and early winter months. According to Birur Mallappa Amrutha et al (2023) When compared to the internal transcribed spacer 2 (ITS2), the mitochondrial cytochrome c oxidase I (COI) was found to be the preferred molecular marker. The interspecific differences between isolates of *R. annulatus* and *R. microplus* was 7.9% based on COI from South India. Furthermore, the *R. microplus* isolates showed greater intraspecific divergence (2.9%) compared to *R. annulatus* (1%), according to COI. The ITS2 sequences were unable to distinguish between *R. annulatus* and *R. microplus*” (Tamura et al., 2021).

Patrician et al. (2024) were studied on “molecular identification of *Rhipicephalus microplus*. Using phylogenetic analysis and the sequencing of the ITS2 region, 12S rRNA, and 16S rRNA genes, the identities of the *R. microplus* tick species were verified”. Hence 18S rRNA and COI markers useful to molecular identification of *R. microplus* species in this study.

Material and Methods

Study areas: This study was conducted in Chh. Sambhajinagar District M.S. India. Selection of cattle which are from different villages in Chhatrapati Sambhajinagar district. Farmers prefer to cattle farming as a side business along with farming. Generally climatic conditions are favourable for cattle but from last few years rising temperature and unpredictable weather patterns all have drastic impact on cattle health.



Image 1: Collection of ticks from various sites in Chh. Sambhajinagar district

Tick collection and identification: The Tick Specimens were collected from the infested cattle by handpick method during of period 2023-24 from various areas of Chh. Sambhajinagar (MS) India. preservation of ectoparasite in 70% alcohol in well stopped glass vials with labelling parasites and Mounted with DPX on cavity slide. Presumptive Identification with help of Understereoscopic microscope. Final Identification will observe under compound microscope according to Keys & description.

Molecular studies

DNA extraction, polymerase chain reaction (PCR), and sequencing:

1. The DNA was extracted by TAKARA NucleoSpin® Tissue Genomic DNA Purification Kit and quality checked on 1% agarose gel electrophoresis. Gel was visualized using UV Transilluminator (Himedia).
2. Fragment of gene COI & 18S rRNA was amplified by 1A and 564R primers.
3. A single discrete PCR amplicon band was observed when resolved on 1.2% Agarose gel.
4. The PCR amplicon was purified to remove contaminants.
5. Forward and reverse DNA sequencing reaction of PCR amplicon was carried out with forward primer and reverse primers using BDTv3.1 Cycle sequencing kit on ABI 3730xl Genetic Analyzer.
6. Consensus sequence of COI & 18S rRNA gene was generated from forward and reverse sequence data using Bioedit software.
7. The COI & 18S rRNA gene sequence was used to carry out BLAST with the database of NCBI Genbank database.
8. Based on maximum identity score and alignments using multiple alignment software program Clustal W. Distance matrix was generated and the phylogenetic tree was constructed using MEGA11.

Results and Discussion

Species confirmation using COI & 18S rRNA marker

Amplification of COI & 18S rRNA

The COI & 18S rRNA region was amplified by using primers (Forward and Reverse as detailed in Table 1). Amplified PCR products were visualized on 1.2% agarose gel.

Table 1. Detail of Polymerase Chain Reaction composition

Component	Component Volume
GoTaqGreen Mastermix	25 μ L
F primer	3 μ L
R primer	3 μ L
Template DNA	6 μ L
Nuclease-Free Water	13 μ L
Total reaction volume	50 μ L

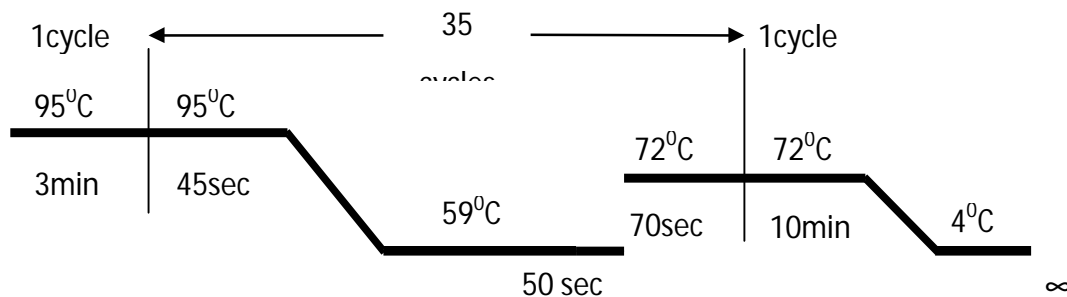


Figure 1. COI & 18S rRNA thermal cycling conditions used for amplification

2. Sequencing

PCR products were processed for cleanup to remove unincorporated nucleotide and residual primers using Exonuclease-

I and Shrimp Alkaline phosphatase enzyme followed by cycle sequencing reaction using BigDye® Terminator v.3.1 Cycle Sequencing Kit (Applied Biosystems, Inc.). For Cycle sequencing same PCR primers were used. The thermal cycler conditions were an initial denaturation of 2 min at 96°C and 35 cycles of 30 sec at 96°C, 15 sec at 55°C,

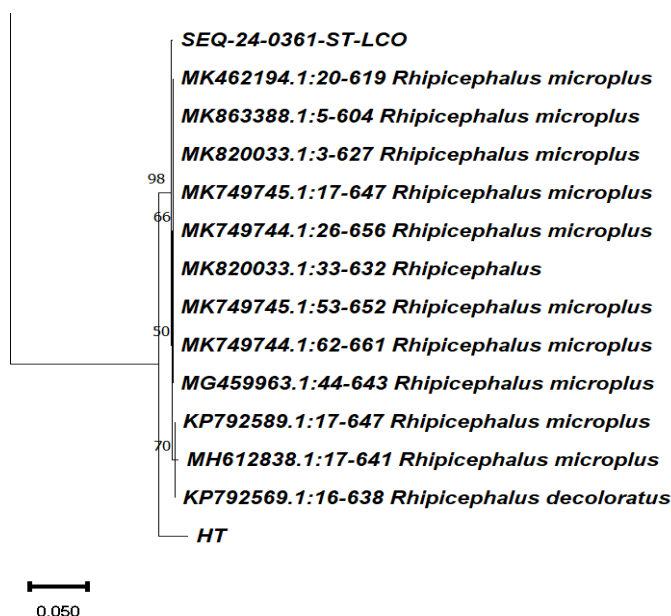
and 4 min at 60°C. The Cycle sequencing is followed by sequencing cleanup by ethanol precipitation followed by dissolving template in HiDi formamide and bidirectionally sequenced in ABI3730 Genetic analyzer.

Table 2. Sample IDs showing Similarity Searches in Sequence Alignment

PCR products were then processed for direct bi-directionally sequencing using ABI PRISM 3730×1 Genetic Analyzer (Applied Biosystems, USA). The resulting DNA sequences were aligned using CLUSTALW in MEGA 11, manually trimmed and edited to obtain complete sequences. The confirmation of species depends on the sequence similarity score.

Homology searches were carried out using the BLAST program against the NCBI GenBank database (<https://blast.ncbi.nlm.nih.gov/Blast.cgi>). Neighbor Joining tree was constructed using MEGA 11 with all positions containing gaps and missing data were included for analysis. Clade supports were calculated based on 1,000 bootstrap samplings.

Fig 2. Search result in BLAST program against the NCBI GenBank



Evolutionary relationship of taxa COI Based

SAMPL EID	Description	MaxScore	TotalScore	QueryCoverage	E-Value	Per Identity	AccessionNo.
HT	<i>Rhipicephalus microplus</i> Isolate 4 cytochrome oxidase subunit I (Cox1) gene partial cds, mitochondrial	1033	1033	97%	0.0	96.20%	KP792589
ST	<i>Rhipicephalus microplus</i> isolate PAK7 cytochrome oxidase subunit 1 (cox1) gene, partial cds; mitochondrial	1098	1098	100%	0.0	99.67%	MK462194

The evolutionary history was inferred using the Neighbor-Joining method. The optimal tree is shown.

The percentage of replicate trees in which the associated taxa clustered together in the bootstrap test

(1000 replicates) are shown above the branches. The tree is drawn to scale, with branch lengths in the same units as those of the evolutionary distances used to infer the phylogenetic tree. The evolutionary distances were computed using the p-distance method, and are in the units of the number of base differences per site. This analysis involved 18 nucleotide sequences.

Codon positions included were 1st+2nd+3rd+Noncoding. All ambiguous positions were removed for each sequence pair (pairwise deletion option). There were a total of 651 positions in the final dataset. Evolutionary analyses were conducted in MEGA11.

Our study utilized a molecular approach, incorporating PCR, DNA sequencing and phylogenetic analysis, to investigate the diversity of ticks.

Conclusion

The results of the present study confirmed the widespread existence of *R. microplus* in Chhatrapati Sambhajinagar district. The result highlighted genetic analysis to confirm true species status of *R. microplus*. Out of two molecular markers, COI was successful in resolving the phylogenetic relationship of *R. microplus*. This study is helpful for researchers to resolve diversity of tick species from genetic analysis.

Disclaimer (Artificial Intelligence)

Authors hereby declare that generative AI technologies such as QuillBot software has been used during the writing or editing of manuscripts for grammar checking and summarizing.

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